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Evaluation of 6 Methods for Aerobic Bacterial Sanitization of Smartphones

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Smartphones are ubiquitous devices that offer a variety of useful applications for human and veterinary medical professionals and the biomedical research community. Smartphones can serve as fomites and potentially transmit pathogens, including bacterial species such as methicillin-resistant *Staphylococcus aureus*. The goal of this study was to evaluate 6 methods to decrease aerobic bacterial colonies on smartphones, including two 254-nm UVC devices, 70% ethanol spray, quaternary ammonium disinfectant spray, sodium hypochlorite-impregnated wipes, and delicate-task wipes. All methods were individually effective at decreasing aerobic bacterial counts after sanitization. In addition, 254-nm UVC devices providing a dose of 60 mJ/cm^2, with UVC bulbs exposing both sides of the smartphone, were an effective nonliquid method for smartphone sanitization.

Portable smart application-based devices have steadily increased in popularity in the United States, with recent surveys suggesting that 72% of adults own a smartphone and 45% own a tablet.[17,18] Portable touchscreen devices offer general applications, such as text messaging, calculators, timers, flashlights, and cameras; medical applications including pharmaceutical formularies, medical calculators, and patient communication; and laboratory-animal specific applications, including electronic medical records, animal census tools, and veterinary pharmaceutical formularies.

Mobile phones can harbor bacterial nosocomial pathogens, including methicillin-resistant *Staphylococcus aureus*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Klebsiella* spp., *Enterobacter* spp., enterococci, and streptococci, among others.[13-15,20,22-24] The few published reports focusing on touchscreen-based smart devices likewise confirm contamination with bacterial pathogens, with one study finding a higher rate of pathogen contamination for smartphones (34.8%) as compared with nonsmartphones (20.5%).[7,11,13]

The potential ability of smartphones to serve as fomites has important implications for laboratory animal facilities. Specifically, the use of contaminated smart devices inside vivaria or procedure rooms poses the risk of exposing research animals to potential pathogens. This risk is especially critical when maintaining SPF genetically modified, immunodeficient, and humanized mouse models, because large commercial mouse vendors exclude bacterial pathogens including *S. aureus* and *P. aeruginosa* from mice housed in their cleanest health-status barrier facilities.[4,9,12,21] Previously, transmission of *Corynebacterium bovis* to a strain of hirsuite immunologically altered mice was attributed to a mobile tablet shared between 2 housing rooms.[5]

Despite the number of publications assessing microbiologic contamination of smartphones, limited published reports evaluate the efficacy of sanitization methods, and those that assess decreases in bacterial colonization after smartphone sanitization evaluate alcohol-impregnated lens wipes, quaternary ammonium disinfectant–detergent, and microfiber cloths.[7,16,20] Additional methods of smartphone disinfection, including commercially available 254-nm UVC smartphone sanitizing devices and bleach-impregnated wipes, have not previously been evaluated, to our knowledge. The goal of this study was to evaluate the efficacy of UV smartphone sanitizing devices as compared with liquid methods of sanitization. We hypothesized that UV light would be more effective than liquid-based methods for sanitization of smartphones.

Materials and Methods

Smartphone inclusion criteria. The characteristics of the smartphones sampled are shown in Figure 1. Smartphones were excluded from the study when physical flaws (for example, a cracked screen) were present and when the smartphone case completely enclosed the phone screen. Smartphone owners were queried regarding whether they sanitized their smartphone regularly, and a minimum of 2 wk since the last sanitization was required prior to sampling. Smartphones that underwent repeated sampling over the course of the project had a minimum intersampling interval of 3 wk. All smartphone sampling procedures were considered exempt by the MIT Committee on the Use of Humans as Experimental Subjects. Written informed consent was obtained for all persons volunteering their smartphones.

Smartphone and UV device sampling. Three separate areas were chosen for sampling and designated as smartphone face, junction, and case. A sterile cotton-tipped applicator (Puritan Medical Products, Guilford, ME) was moistened in tryptic soy broth (Becton Dickinson, Sparks, MD) and swabbed over the smartphone face, smartphone–case junction, and the sides and back of the case (Figure 2 A). Each swab was rolled down and up the center of a plate containing trypticase soy agar with 5% sheep blood (Figure 2 B). A disposable 10-µL inoculating loop (Greiner Bio-One, Monroe, NC) was used to repeatedly spread colonies across the plate, perpendicular to the initial swab (Figure 2 B). Each area was sampled before and after sanitization. The smartphone without a case was sampled by using 2 swabs, encompassing the smartphone face and combined smartphone sides and back. The same persons performed all swab procedures and initial inoculation of agar plates (MTL) and dispersion of the swab inoculum (CMM). As a control, UVC devices were sampled by moistening a cotton-tipped applicator in tryptic soy

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	Description	*n*
Smartphone		
Apple	iPhone 4/4S	3
	iPhone 5S/5c/SE	11
	iPhone 6/6S/7	6
	iPhone 6S+	1
Motorola	Droid Mini	2
	Moto G	1
Screen protector		
None	Not applicable	18
Adhesive	Plastic	4
	Glass	2
Case		
None	Not applicable	1
Bumper	Soft silicone	2
	Hard silicone	2
	Soft plastic	1
	Hard plastic	8
	Hard plastic + hard silicone	5
	Hard plastic + soft silicone	3
Wallet	Leather	1
	Hard plastic + canvas	1

Figure 1. Characteristics of smartphones sampled (n = 24).

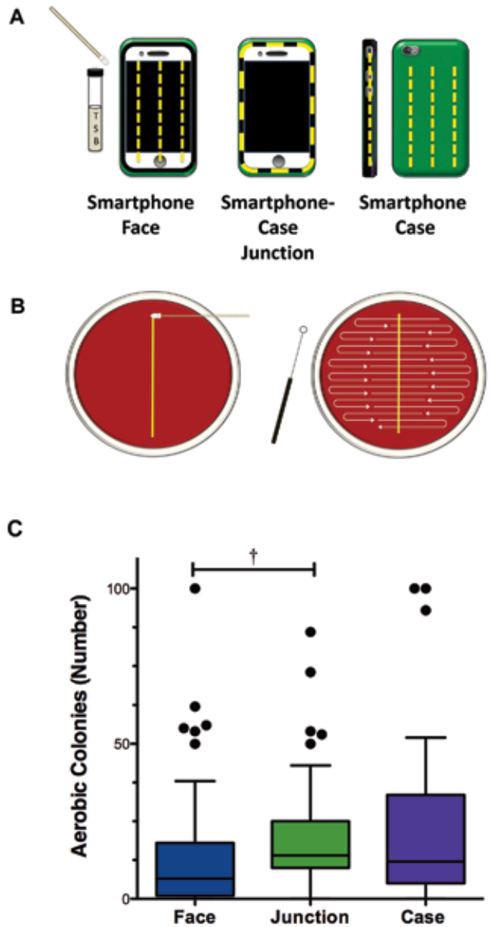

Figure 2. Smartphone sampling procedure and contamination measured by using aerobic bacterial counts at each sampling site. (A) A sterile cotton-tipped applicator moistened with tryptic soy broth (TSB) was used to sample the front of the smartphone face, smartphone-case junction, and sides and back of the smartphone case as designated by yellow dashed lines. (B) Swabs from smartphone sampling were plated down the center of a tryptic soy agar plate with 5% sheep blood, and the initial swab inoculum was spread in perpendicular manner by using a 10-µL disposable inoculating loop. (C) The smartphone face was significantly less contaminated than the phone-case junction. Tukey box and whiskers plot; Kruskal–Wallis with Dunn multiple comparison test, $P = 0.0046$. †, $P \leq 0.01$.

broth and swabbing the surface where the smartphone would be placed. The swabs were inoculated onto the first quadrant of trypticase soy agar with 5% sheep blood, and a sterile inoculating loop was used to streak for colony isolation. Plates were incubated overnight at 37 °C with 5% CO_2.

Sanitization methods. All smartphones were left inside their cases during sanitization. Sanitization methods included 2 commercially available smartphone UVC-sanitizing devices (PS300, PhoneSoap, Provo, UT; FBM120, Flashbox mini, ClorDiSys Solutions, Branchburg, NJ; Figure 3), 70% ethanol spray, 0.55% sodium hypochlorite wipes (Bleach Germicidal Wipes, Clorox Healthcare, Oakland, CA), quaternary ammonium disinfectant spray (Quatricide PV, Pharmacal, Naugatuck, CT), and cleaning with a delicate-task wipe (KimWipe, Kimberly-Clark Professional, Roswell, GA). For the PS300, the smartphone was placed inside the device, screen-side up, and the lid was closed to activate the UV light, according to the manufacturer's instructions, for a set sanitizing time of 5 min. For the FBM120, the smartphone was placed screen-side up on the glass shelf set on the lowest height. The light mode was set to 'all' (for both top and bottom UV-light activation), and the time knob switched past 2 min while an independent timer was set for 2 min. For the spray sanitization methods of 70% ethanol and quaternary ammonium disinfectant, smartphones were sprayed twice (front and back) by using a squirt bottle at a distance of approximately 12 in. to mimic a realistic disinfection procedure. Smartphones were wiped immediately, front and combined back and sides, by using a clean paper towel in a single downward motion and were placed on top of a clean paper towel for sampling. For sanitization by using bleach wipes, smartphones were wiped front, sides, and back by using one wipe per smartphone and then allowed to dry completely on a clean paper towel prior to sampling. For sanitization by using a delicate-task wipe, the smartphone was wiped on the front, sides, and back by using a single wipe and placed on a clean paper towel for sampling. We evaluated 7 smartphones per method, except for the PS300

	PS300	FBM120
Recommended sanitizing time (min)	5	2[a]
Number of UVC bulbs (location)	2	2
UVC power per Bulb (W)	1[b]	8[b]
Irradiance ($\mu W/cm^2$)[c]	200	500
UVC dose (mJ/cm^2)[d]	60	60

[a] Manufacturer states a 99% reduction of methicillin-resistant *S. aureus* in 10 s and of *Clostridium difficile* spores in 2 min; we therefore chose 2 min as the sanitizing time.
[b] 1 bulb each at top and bottom
[c] Information provided by the manufacturer
[e] Calculated by multiplying total irradiance by sanitization time

Figure 3. Characteristics of 254-nm UVC sanitizing devices.

(*n* = 9). Two additional smartphones were tested by using the PS300 method because 3 phones tested had presanitization bacterial colony counts of 0 for the smartphone face sampling site.

Colony enumeration. After overnight incubation at 37 °C with 5% CO_2, aerobic colony counts were enumerated for each plate. For the purposes of statistical analysis, plates with colonies too numerous to count were designated with a count of 100. The same person performed colony enumeration (CMM) for all samples.

Statistical analysis. Data were analyzed by using Prism 5.0 (GraphPad Software, La Jolla, CA). We compared the number of aerobic colonies prior to sanitization to determine the most likely contamination site on smartphones by using the Kruskal–Wallis test with Dunn multiple-comparison tests. The effect of a screen protector on number of aerobic colonies prior to sanitization of the smartphone face was evaluated by using the Mann–Whitney test. To evaluate each method of sanitization, we compared the numbers of aerobic colonies before and after sanitization by using Wilcoxon signed-rank tests. To normalize efficacy relative to the presanitization number of bacterial colonies and to compare efficacy between sanitization methods, percentage reduction in aerobic colony count was calculated by using the following equation:

$$\%\ \text{reduction} = 100\% \times \frac{(\text{no. of colonies before sanitation} - \text{no. of colonies after sanitation})}{\text{no. of colonies before sanitation}}$$

Percentage reduction in aerobic colony count was evaluated for the combined smartphone face, smartphone–case junction, and smartphone case, unless otherwise indicated. Comparison in percentage reduction across different sanitization methods was evaluated by using the Kruskal–Wallis test with the Dunn multiple-comparison test. Smartphones with a presanitization colony count of 0 were excluded from percentage reduction analysis. For the smartphone that lacked a case, the back and sides of the smartphone were treated as the case sampling area. Categorical comparisons for a reduction in aerobic colony count to 0 were evaluated by using Pearson χ^2 analysis among sanitization methods. The significance level for all tests was set as an α value of 0.05.

A simple Bayesian estimation model was built to model the uncertainty surrounding the percentage reduction for all sampling sites, given the available data. Briefly, the presanitization and postsanitization colony counts were modeled by using Poisson likelihood, with a DiscreteUniform (0, 1000) prior placed on the Poisson rate parameter. The notation is as follows:

$$\mu_{pre} \sim \text{DiscreteUniform}(0, 1000)$$

$$\text{colony count}_{pre} \sim \text{Poission}(\mu_{pre})$$

$$\mu_{post} \sim \text{DiscreteUniform}(0, 1000)$$

$$\text{colony count}_{post} \sim \text{Poission}(\mu_{post})$$

Percentage reduction, δ_p, was then computed deterministically from the estimated µ posterior distributions:

$$\delta_p = \frac{\mu_{pre} - \mu_{post}}{\mu_{pre}} \times 100\%$$

The Bayesian estimation model was implemented in PyMC3, version 3.0 (https://github.com/pymc-devs/pymc3/archive/v3.0.zip) in the Python programming language (version 3.5). Notebooks are available on GitHub (https://github.com/ericmjl/mia-stats/blob/master/sterilization/sterilization.ipynb) and are archived on Zenodo (DOI: http://doi.org/10.5281/zenodo.275624).

Results

Experiments were designed to mimic realistic conditions; consequently, smartphones were used 'as-is' and sanitized in a manner approximating a real-life scenario. We first determined overall colonization density at each smartphone sampling site and the efficacy of each sanitization method individually before comparing sanitization methods.

Contamination by site. The median (range) for the number of aerobic bacterial colony-forming units present on the smartphone face, smartphone–case junction, and smartphone case prior to sanitization were 6.50 (0 to 100), 14.0 (0 to 86), and 12.0 (0 to 100) colonies, respectively, with the smartphone face having significantly fewer colonies present than the junction ($P = 0.0046$, Kruskal–Wallis; Figure 2 C). The number of aerobic colonies before sanitization did not differ between smartphone faces with a screen protector compared with those without a screen protector ($P = 0.6827$; Mann–Whitney test). The number of aerobic colonies present on smartphone cases before sanitization did not differ between the 3 most common case types—hard plastic, hard plastic–hard silicone, and hard silicone ($P = 0.0532$, Kruskal–Wallis). All cultures from the UVC devices were negative for aerobic growth.

Individual sanitization efficacy. The number of colonies present before sanitization did not differ between methods tested ($P = 0.5895$, Kruskal–Wallis with Dunn multiple-comparison test). Each method tested was individually effective at reducing the number of aerobic colonies after sanitization (Figure 4).

Percentage reduction in aerobic colony count. For all smartphone sampling sites combined, the FBM120 device achieved a significantly ($P < 0.05$) higher percentage reduction in colony count than all other sanitization methods except the PS300 device (Figure 5 A). The PS300 sanitization method showed a higher percentage reduction as compared with quaternary ammonium disinfectant and delicate-task wipes (Figure 5 A). When analyzed by sampling location, Kruskal–Wallis analysis found a significant difference in the percentage reduction at the smartphone face between sanitization methods ($P = 0.0462$); however, posthoc testing did not identify pairwise differences (Figure 5 B). For the smartphone–case junction, the FBM120 and PS300 devices showed higher percentage reductions in colony count compared with the delicate-task wipe (Figure 5 C). For

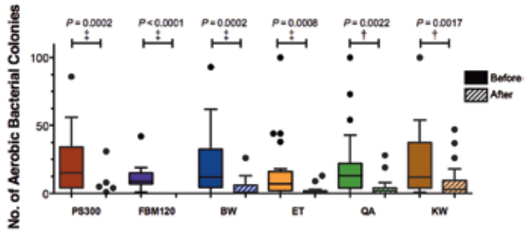

Figure 4. Efficacy of sanitization methods in reducing aerobic bacterial colony count. Tukey box and whiskers plot. Wilcoxon signed-rank test, with *P* values indicated above each treatment. Solid bars, presanitization colony counts; striped bars, postsanitization colony counts; †, $P < 0.01$; ‡, $P < 0.001$; BW, bleach wipe; ET, 70% ethanol; QA, quaternary ammonium disinfectant spray; KW, delicate-task wipe.

the smartphone case, the FBM120 showed a higher percentage reduction in colony count than quaternary ammonium disinfectant (Figure 5 D). The Bayesian estimation model supported these results, revealing that the FBM120 and PS300 devices showed the most consistent percentage reduction in colony count with the narrowest 95% credible intervals (Figure 6).

Efficacy in reducing aerobic bacterial colony count to 0. We performed a χ^2 test to evaluate the relationship between sanitization method and efficacy in reducing the aerobic bacterial colony count to 0 (that is, aerobic bacterial sterilization). The observed χ^2 was 40.82 with 5 degrees of freedom and $P < 0.0001$, allowing rejection of the null hypothesis that all sanitization methods were equally likely to reduce the aerobic colony count to 0 (Figure 7). All 7 smartphones sanitized with the FBM120 demonstrated 100% reduction in aerobic bacterial colonies to 0 at all 3 sampling sites after sanitization.

Discussion

In addition to efficacy in decreasing bacterial burden, several factors should be considered when establishing appropriate biosecurity protocols for smartphones and smart devices in the healthcare and preclinical research settings. To encourage compliance, sanitization methods should work rapidly (ideally, in the time it takes to wash hands or apply or remove personal protective equipment) and not damage mobile devices. Because of the risks of smartphone damage with exposure to liquid disinfectants, UVC sanitization methods are potentially superior to disinfection with bleach, ethanol, or quaternary ammonium solutions. None of the liquid disinfection methods used damaged the sampled smartphones in the current; however, we cannot comment on device impairment after repeated liquid disinfectant exposures. A previous study recovered pathogenic bacteria on 44 of 53 (83%) cell phones in a hospital environment; immediately after sanitization with an isopropanol-impregnated lens wipe, 4 of the 53 (8%) cell phones remained culture-positive for pathogenic bacteria.[20] A second study found no bacterial growth on touchscreens after cleaning with an ethanol–isopropanol-impregnated lens wipe in 35% of phones ($n = 20$).[7] Our results for sanitization with 70% ethanol are intermediate to these findings, with 39% of sampling sites (7 of 18 sites among 7 phones) showing no aerobic growth immediately after sanitization (Figure 7). Another publication reported that a 0.25% concentration of a detergent–disinfectant combination containing N-(3-aminopropyl)-N-dodecylpropane-1 and didecyldimethylammonium chloride reduced colony counts by approximately 50%; however only 25% (13 of 52) phones were free of colony growth following decontamination procedures.[16] Our results for quaternary ammonium disinfection are in agreement, with 28% of sampling sites (5 of 18 sites among 7 phones) showing no aerobic growth after sanitization.

Our results indicated that the smartphone–case junction had significantly more contamination than the smartphone face. We hypothesize that this situation results from the persistence of debris and bacteria in the crevice between the smartphone and the case (Figure 2 C). UV sanitization methods had a smaller interquartile range for percentage reduction in aerobic colony counts for the smartphone–case junction as compared with liquid and wipe methods (Figure 5 C). We found it surprising that the case did not have a higher presanitization bacterial burden; this result may reflect the sample size and lack of standardization of case material and styles.

Physically wiping the phone is a variably effective method for decontamination when no other sanitization options are available. The delicate-task wipe was far less effective that UV-based methods when sanitizing the smartphone–case junction (Figure 5 C). A previous report found that 25% of touchscreens ($n = 20$) did not show any bacterial growth after cleaning with a new, dry, microfabric cloth.[7] Our results found that the delicate-task wipe was effective at reducing bacterial burden by 55% to 70% overall, with no aerobic growth on 29% of sampling sites (6 of 21 sites among 7 phones; Figures 6 and 7).

During analysis of the results, there were several instances where the percentage reduction in colony count was negative, that is, more bacterial colonies were present on the smartphone after sanitization than before. Because no growth occurred after aerobic culture of the interior of the UV devices, we believe that increased postsanitization bacterial counts likely resulted from inconsistent speed when obtaining the swab samples. Specifically, a slower swabbing rate increases contact time between the phone and swab, allowing for more bacteria to be sampled. We also cannot rule out UVC bulb failure in the PS300, because the device design prohibits the UVC light from turning on when the device is open. One PS300 sanitization procedure yielded a marked increase in the case postsanitization case colony count, yet, the postsanitization smartphone face and phone–case junction colony counts decreased, suggesting either bulb malfunction or error during the postsanitization swabbing procedure. Another possible source for increased bacterial counts after spray sanitization methods with quaternary ammonium disinfectant or 70% ethanol are contamination of the spray nozzle or paper towel used for sanitization. Paper towels were removed directly from a shared paper towel dispenser to mimic everyday conditions.

Between the 2 commercially available 254-nm UVC devices evaluated in this study, we found that the FBM120 was superior to the PS300 for consistency in reducing the bacterial burden to 0 at all sampling sites and required only a 2-min sanitizing period. UVC kills cells through the induction of pyrimidine dimers in DNA, thus disrupting the DNA replication process.[8] The effectiveness of UVC sterilization is dependent on the dose, which is defined as the amount of UV energy (mJ) per unit area (cm^2) and sometimes expressed as irradiance ($J/s/cm^2$ or W/cm^2). Doses of 15 mJ/cm^2 are able to achieve 3- to 4-log reductions in *Klebsiella pneumoniae*, *Citrobacter* spp., *Escherichia coli*, *Salmonella typhi*, *Shigella sonnei*, *Staphylococcus aureus*, and *Enterococcus* (*Streptococcus*) *faecalis*; however, higher doses are required for rotavirus and poliovirus (>30 mJ/cm^2) and *Bacillus subtilis* spores (>60 mJ/cm^2).[2,10] The PS300 device contains two 1-W, 254-nm bulbs, 1 each located on the bottom and lid of the device and providing an output of 200 $\mu W/cm^2$ each, at the distance to the phone (according to the manufacturer). This setup achieves a UVC dose of 12 mJ/cm^2 for a 1-min exposure

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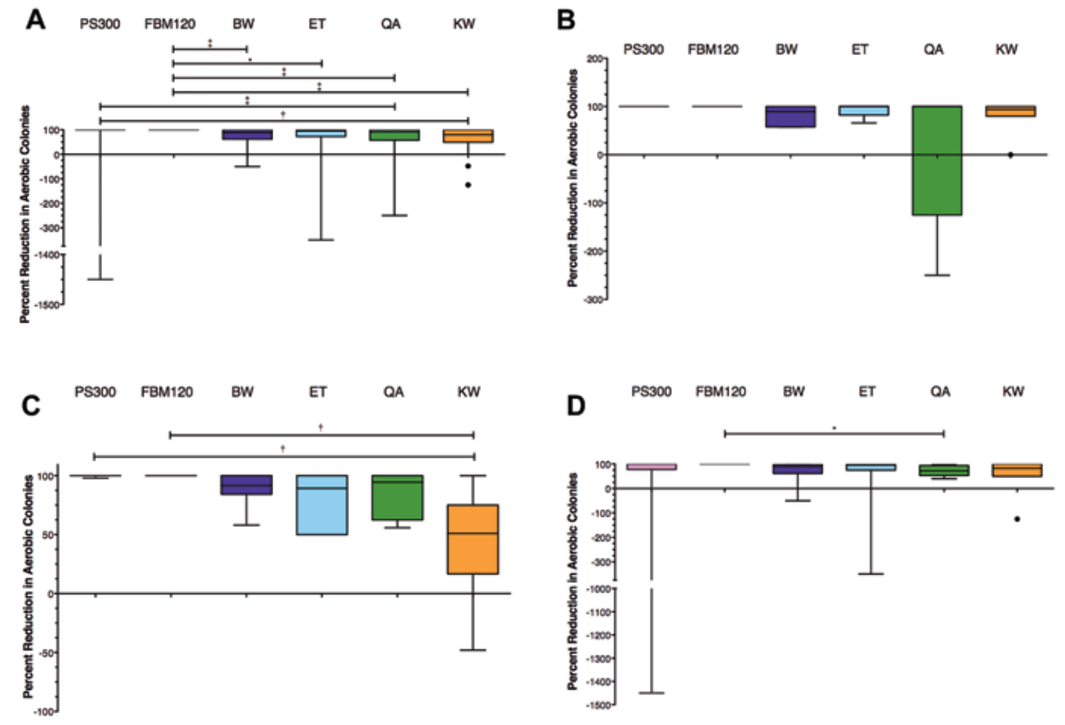

Figure 5. Percentage reductions in aerobic bacterial count for all sampling sites. Tukey box and whiskers plots. In this figure, maximum sanitization effectiveness is represented by100% reduction. (A) Combined colony count for all sampling sites. Kruskal–Wallis with Dunn multiple-comparison test, $P < 0.0001$; posthoc testing: *, $P \leq 0.05$; †, $P \leq 0.01$; ‡, $P \leq 0.001$. (B) Smartphone face ($P = 0.0462$). (C) Smartphone–case junction ($P = 0.0011$). (D) Smartphone case ($P = 0.0322$); posthoc testing: *, $P \leq 0.05$; †, $P \leq 0.01$.

and a total experimental dose of 60 mJ/cm^2 for the 5-min sanitization. The FBM120 device contains two 8-W bulbs, 1 each located on the ceiling and the floor of the sanitizing chamber, and provides a total output of 500 $\mu W/cm^2$ at a 3-in. distance, equal to 30 mJ/cm^2/min (according to the manufacturer) and a total experimental dose of 60 mJ/cm^2 for the 2-min sanitization period.

Our results also indicated that the FBM120 device was the most effective and most consistent method for sanitizing smartphones. Although the PS300 device was highly effective in sanitizing the smartphone face and smartphone–case junction, it did not achieve 100% reduction in aerobic colonies to 0 after sanitization of the smartphone case.

We recognize that a limitation of this study is that we did not perform identification procedures on the bacterial colonies isolated from the smartphones. We did observe that the most common colony morphologies noted were consistent with *Staphylococcus*, *Streptococcus*, and *Bacillus* spp., and we acknowledge previous studies characterizing nonpathogenic and potentially pathogenic bacteria isolated from personal mobile devices.[1,3,13,14,20,23,24] We also recognize that constraining the maximal number of colonies enumerated to 100 for statistical analysis potentially underestimates the calculation for percentage reduction in aerobic colony counts. The number of presanitization smartphone sampling sites designated as 'too many to count' represented a minority of our samples (3 of 131 samples; that is, 2% of swabs). In addition, reduction of aerobic bacteria count to 0 is unaffected by this

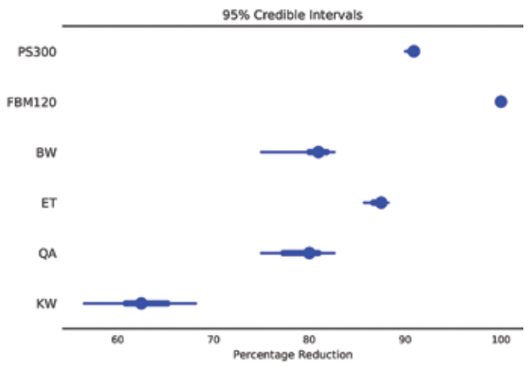

Figure 6. Bayesian estimation model of sanitization efficacy for all smartphone sampling sites combined. The posterior distribution is summarized in blue: the dot represents the median, the thick blue line indicates the interquartile range, and the thin blue line indicates the width of the 95% credible interval probability mass.

constraint and is arguably the most important parameter when evaluating sanitization efficacy.

In the preclinical and laboratory animal research settings, other highly resistant pathogens of interest to exclude from rodent colonies include mouse parvovirus and pinworms, especially those of the genus *Syphacia*. Previous studies

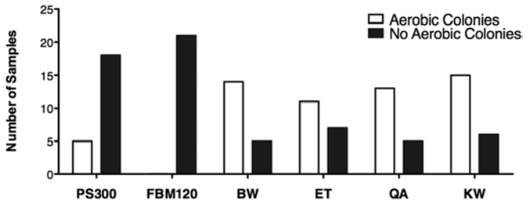

Figure 7. χ^2 analysis of the efficacy in reducing aerobic bacterial colony count to 0 for all sampling sites combined. $\chi^2 = 40.82$; degrees of freedom, 5; $P < 0.0001$.

suggest that 254-nm UVC light is effective at inactivating porcine parvovirus[19] as well as preventing hatching of *Syphacia muris* ova,[6] although future studies should investigate the effectiveness of small, portable 254-nm UVC devices on these agents.

With increasing use of portable mobile devices in research and human and veterinary healthcare settings, appropriate biosecurity protocols must be established to prevent fomite transmission of bacterial pathogens. Our results indicate that UVC-based sanitization methods are effective in reducing bacterial burden, but not all devices are equivalent in their abilities to reduce aerobic bacterial colonization to 0. Our evaluation of 2 commercially available 254-nm UVC sanitizing devices suggests that sanitizing devices providing a total UVC dose of approximately 60 mJ/cm^2 and UVC bulb exposure on both sides of the smartphone are effective in sanitizing smartphones and reducing the aerobic bacterial count to 0.

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Time Required to Achieve a Given Log Reduction using the Flashbox mini™

	1-Log (90%)	2-Log (99%)	3-Log (99.9%)	4-Log (99.99%)	Reference
Spore					
Bacillus anthracis spores - Anthrax spores	1 min	2 min			Light Sources Inc. 2014
Bacillus subtilis ATCC6633	1 min	1.5 min	2 min	3 min	Mamane-Gravetz and Linden 2004
Closttridium difficile spores			2 min		Antimicrobial Test Laboratories 2015
Bacterium					
Bacillus anthracis - Anthrax	.5 min	.5 min			Light Sources Inc. 2014
Campylobacter jejuni ATCC 43429	.5 min	.5 min	.5 min	.5 min	Wilson et al. 1992
Clostridium tetani	.5 min	1 min			Light Sources Inc. 2014
Corynebacterium diphtheriae	.5 min	.5 min			Light Sources Inc. 2014
Escherichia coli	.5 min	.5 min			Light Sources Inc. 2014
Escherichia coli O157:H7	.5 min	.5 min	.5 min	.5 min	Tosa and Hirata 1999
Klebsiella pneumoniae	.5 min	.5 min	1 min	1 min	Giese and Darby 2000
Legionella pneumophila	.5 min	.5 min	.5 min	.5 min	Oguma et al. 2004
Mycobacterium tuberculosis	.5 min	.5 min			Light Sources Inc. 2014
Pseudomonas aeruginosa	.5 min	.5 min			Light Sources Inc. 2014
Salmonella enteritidis	.5 min	.5 min	.5 min	.5 min	Tosa and Hirata 1998
Salmonella typhosa - Typhoid fever	.5 min	.5 min			Light Sources Inc. 2014
Shigella dyseteriae - Dysentery	.5 min	.5 min			Light Sources Inc. 2014
Staphylococcus aureus ATCC25923	.5 min	.5 min	.5 min	.5 min	Chang et al. 1985
Vibrio comma - Cholera	.5 min	.5 min			Light Sources Inc. 2014
Molds					
Aspergillius flavus	2 min	3.5 min			Light Sources Inc. 2014
Aspergillius niger	4.5 min	11 min			Light Sources Inc. 2014
Mucor racemosus A & B	1 min	1.5 min			Light Sources Inc. 2014
Viruses					
Adenovirus type 15	1.5 min	3 min	4.5 min	5.5 min	Thompson et al. 2003
Adenovirus type 2	1 min	1.5 min	3 min	4 min	Shin et al. 2005
Bacteriopfage - E. Coli	.5 min	.5 min			Light Sources Inc. 2014
Calicivirus canine	.5 min	.5 min	1 min	1 min	Husman et al. 2004
Calicivirus feline	.5 min	1 min	1 min		Husman et al. 2004
Coxsackievirus B5	.5 min	1 min	1 min	1.5 min	Gerba et al. 2002
Hepatitis A	.5 min	.5 min	.5 min	1 min	Wiedenmann et al. 1993
Hepatitis A HM175	.5 min	.5 min	1 min	1 min	Wilson et al. 1992
Influenza	.5 min	.5 min			Light Sources Inc. 2014
Norovirus			1 min		Lee et al. 2008
Poliovirus 1	.5 min	1 min	1 min	1.5 min	Gerba et al. 2002
Staphylococcus aureus phage A 994	.5 min	1 min	1 min	1.5 min	Sommer et al. 1989

This chart describes the required dosage time necessary to achieve a given log reduction of that particular organism, based on published data. Times are rounded up to the nearest half minute. The chart can be used to determine the necessary length of UV-C exposure time is needed to get the disinfection level desired.

Flashbox mini™

Description:

The Flashbox mini™ UV-C Disinfection Chamber is an easily transportable, small chamber designed for use in any healthcare, pharmaceutical, manufacturing, laboratory, or research setting. It is used to provide a rapid and highly effective method to disinfect tablet computers, phones, remote controls, miscellaneous electronics, instruments, and components to reduce the transfer of dangerous organisms. The Flashbox mini™ also offers a way to disinfect components without removing them from the room, which helps minimize the chance for cross-contamination.

The Flashbox mini™ contains 1 quartz glass shelf to support the item(s) being disinfected. It simply plugs into any wall outlet. The disinfection chamber produces an efficient UV-C output of approximately 500 µw/cm^2 to get a calculated 99.99% reduction of MRSA in 1 minute and a 99% *Clostridium difficile* spores in 2 minutes.

Specifications:

Usable Space for items: 3.25″H x 8.5″D x 9.7″W

Overall Dimensions: 8.25″H x 11″D x 14.5″W
Power: 115 VAC, 60 Hz, 2 Amps

UV-C Output: 30 mJ/cm^2 per minute (500 µw/cm^2) at a 3″ distance.

Features:

Efficacy:

- The Flashbox mini™ contains 2 protected UV-C bulbs, one on the top and one on the bottom, to provide increased disinfection coverage of items placed inside the chamber.
- At the furthest point from the bulbs, the FLASHBOX-mini provides over 500 µW/cm^2 of UV-C intensity. This intensity correlates to a 30 mJ/cm^2 UV-C dosage during a one-minute exposure.
- The Flashbox mini's™ UV-C output was validated using two independent UV-C Sensors, the Solar Light Company's PMA1122 Germicidal UV-C Sensor and the General® UV512C Digital UV-C Meter.

Operation:

- Easily operated with minimal training.
- No chemicals to store and handle.
- Simple manual timer to set disinfection time.
- The Flashbox mini™ has a semi-transparent door, allowing visual confirmation that the unit is working properly.

Safety:

- The door contains a safety switch which turns the unit off if the door is opened during an exposure.
- The glass door blocks UV-C wavelengths from passing through, such that it is safe to look through the glass while the unit is running.

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